Immunophenotyping and extracellular matrix remodeling in pulmonary and extrapulmonary sarcoidosis*,**


Abstract

Objective: To investigate the significance of cellular immune markers, as well as that of collagen and elastic components of the extracellular matrix, within granulomatous structures in biopsies of patients with pulmonary or extrapulmonary sarcoidosis. Methods: We carried out qualitative and quantitative evaluations of inflammatory cells, collagen fibers, and elastic fibers in granulomatous structures in surgical biopsies of 40 patients with pulmonary and extrapulmonary sarcoidosis using histomorphometry, immunohistochemistry, picrosirius red staining, and Weigert’s resorcin-fuchsin staining. Results: The extrapulmonary tissue biopsies presented significantly higher densities of lymphocytes, macrophages, and neutrophils than did the lung tissue biopsies. Pulmonary granulomas showed a significantly higher number of collagen fibers and a lower density of elastic fibers than did extrapulmonary granulomas. The amount of macrophages in the lung samples correlated with FVC (p < 0.05), whereas the amount of CD3+, CD4+, and CD8+ lymphocytes correlated with the FEV1/FVC ratio and VC. There were inverse correlations between TLC and the CD1a+ cell count (p < 0.05), as well as between DLCO and collagen/elastic fiber density (r = −0.90; p = 0.04). Conclusions: Immunophenotyping and remodeling both showed differences between pulmonary and extrapulmonary sarcoidosis in terms of the characteristics of the biopsy samples. These differences correlated with the clinical and spirometric data obtained for the patients, suggesting that two different pathways are involved in the mechanism of antigen clearance, which was more effective in the lungs and lymph nodes.

Keywords: Sarcoidosis; Granulomatous disease, chronic; Extracellular matrix; Immunophenotyping; Respiratory function tests.

Resumo

Objetivo: Investigar o significado de marcadores de imunidade celular e de componentes elásticos/collágeno da matriz extracelular em estruturas granulomatosas em biópsias de pacientes com sarcoidose pulmonar ou extrapulmonar. Métodos: Determinações qualitativas e quantitativas de células inflamatórias, de fibras de colágeno e de fibras elásticas em estruturas granulomatosas em biópsias cirúrgicas de 40 pacientes com sarcoidose pulmonar e extrapulmonar foram realizadas por histomorfometria, imuno-histoquímica, e técnicas de coloração com picrosirius e resorcina-fuscina de Weigert. Resultados: A densidade de linfócitos, macrófagos e neutrófilos nas biópsias extrapulmonares foi significativamente maior do que nas biópsias pulmonares. Os granulomas pulmonares apresentaram uma quantidade significativamente maior de fibras de colágeno e menor densidade de fibras elásticas que os granulomas extrapulmonares. A quantidade de macrófagos nos granulomas pulmonares correlacionou-se com CVF (p < 0,05), ao passo que as quantidades de linfócitos CD3+, CD4+ e CD8+ correlacionaram-se com a relação VEF1/CVF e com CV. Houve correlações negativas entre CPT e contagem de células CD1a+ (p < 0,05) e entre DLCO e densidade de fibras elásticas/collágenas (r = −0,90; p = 0,04). Conclusões: A imunofenotipagem e o remodelamento apresentaram características diferentes nas biópsias dos pacientes com sarcoidose pulmonar e extrapulmonar. Essas diferenças correlacionaram-se com os dados clínicos e espirométricos dos pacientes, sugerindo que há duas vias envolvidas no mecanismo de depuração de antígenos, que foi mais eficaz nos pulmões e linfonodos.

Descritores: Sarcoidose; Doença granulomatosa crônica; Matriz extracelular; Imunofenotipagem; Testes de função respiratória.

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Introduction

Sarcoidosis is a systemic granulomatous disease of unknown etiology. The diagnosis is based on clinical, radiographic, and pathological findings. Most patients generally have favorable clinical courses.\(^{10}\) Although pulmonary and extrapulmonary parenchymal involvement improves spontaneously in most patients, progression to fibrosis, leading to permanent functional impairment, occurs in 20–25%, and death occurs in 5–10%.\(^{1,2}3\) In this context, a large number of clinical, physiological, and radiographic parameters have been investigated in order to evaluate the outcome of sarcoidosis.\(^{1,4\text{-}8}\)

The disease activity is marked by the process of extracellular matrix (ECM) remodeling—which involves the balance between immune cells and tissue remodeling\(^{9,10}\)—the impact of histopathological staging on the prognosis of sarcoidosis having never been investigated.

Because cellular immune phenomena and ECM remodeling have been shown to be promising prognostic markers in diffuse interstitial lung disease,\(^{11-14}\) we hypothesized that the two have a similar impact on the prognosis of sarcoidosis. We reviewed medical records and pathology reports over a period of 10 years in order to investigate the significance of cellular immune markers, as well as that of collagen and elastic components of the ECM, within the granulomatous structures found in biopsies of patients with pulmonary or extrapulmonary sarcoidosis.\(^{15}\)

Methods

The study group comprised 40 patients who had been diagnosed with sarcoidosis on the basis of clinical and histological criteria.\(^{1}\) The tissue samples examined included pulmonary parenchymal and bronchial biopsy specimens (in 25 patients), mediastinal or peripheral lymph nodes (in 8 patients), liver (in 4 patients), and skin (in 3 patients).

The medical records were reviewed, and the date of the biopsy was used as the date of diagnosis. Information regarding patient gender, patient age, and the treatment given was collected. The patients with pulmonary sarcoidosis were in the early stage of the disease and underwent lung biopsy as part of their initial clinical evaluation. Cardiac parameters were normal in those patients, and chest HRCT scans (1.0- or 1.5-mm-thick slices) were taken at 1-cm intervals while the patients took deep breaths in the supine position; for the last 10 cm of the caudal part of the lungs, the scans were taken at 2–3 cm intervals, with the patients in the prone position. Two thoracic radiologists prospectively and independently examined all of the lobes on the HRCT scans for micronodules, opacity, and lymphadenopathy (Table 1).

For all of the patients with pulmonary sarcoidosis, we collected data on the history of smoking and pulmonary function parameters (FVC, FEV\(_1\), FEV\(_1\)/FVC ratio, RV, TLC, and single-breath DLCO). We also collected the scores on a six-point dyspnea scale routinely applied to patients treated at our institution, the scores ranging from 0 (no dyspnea) to 6 (very severe dyspnea).\(^{16}\)

Staining of collagen fibers was performed with a 0.2% solution of Sirius red (Direct Red 80, CI 35780; Aldrich, Milwaukee, WI, USA) dissolved in aqueous saturated picric acid. This dye has been widely used for staining collagen in histological specimens and allows the quantitative analysis of collagen in paraffin sections.\(^{11,17}\) The enhancement of collagen birefringence by picrosirius red staining and polarized light microscopy is specific for all collagen structures consisting of aggregates of oriented molecules. Elastic fiber staining was performed with the Weigert’s resorcin-fuchsin method, after oxidation.\(^{18}\) This method allows selective identification of the three types of elastic fibers (oxytalan, elaunin, and fully developed elastic fibers).

The primary antibodies used in the present study were mouse monoclonal antibodies (Dako, Glostrup, Denmark; for all):

- mouse anti-human CD68, clone KP1 (M0814; dilution, 1:3,200), a marker for macrophages/histiocytes
- mouse anti-human neutrophil elastase, clone NP57 (M0752; dilution, 1:800)
- anti-human CD3 (dilution, 1:600), CD4 (dilution, 1:400), CD8 (dilution, 1:100), CD20 (dilution, 1:600), CD1a (dilution, 1:20), and S100 (dilution, 1:600), all of which are markers for T and dendritic cells

For the tests, the streptavidin-biotin complex method was used.\(^{14,19}\)

Immune cells were quantified by a conventional stereologial method, i.e., the point sampled intercept method.\(^{20}\) At a magnification of x400, 8–10 granulomatous structures were evaluated by
systematic point counting, an eyepiece micrometer and a sampling grid with 100 points and 50 lines having been used in order to count the number of points overlying positively stained cells.

Collagen and elastic fibers were quantified by image analysis. The image analysis system consisted of an Olympus camera coupled to an Olympus microscope (Olympus Optical, Tokyo, Japan), which transmitted the images to a computer equipped with a Pentium 1,330-MHz processor (Intel Corporation, Santa Clara, CA, USA) and a monitor (LG Electronics Brasil, Manaus, Brazil) by means of a digitizing system (Oculus TCX; Coreco Inc., St. Laurent, Quebec, Canada). The images were processed using the program Image Pro-Plus, version 6.0 (Media Cybernetics Inc., Bethesda, MD, USA). For each tissue specimen, a range of 8-10 granulomatous structures were analyzed at ×400 magnification. The density of collagen and elastic fibers was measured and expressed as a ratio between the quantity of fibers and the total area studied. The final results express the area occupied by the various collagen and elastic fibers in relation to the total area.

One-way ANOVA was used in order to analyze the variation in the mean number of immune cells, collagen fibers, and elastic fibers, as well as the distribution of immune cells, collagen fibers, and elastic fibers within granulomas in the various organ tissues. The means were compared, a priori, by Levene’s test for homogeneity of variance, followed by post hoc analysis with the Bonferroni test for multiple comparisons (in case of homogeneity of variance) or Dunnett’s T3 test (in case of heterogeneity of variance). In addition, a paired t-test and the general linear model were used in order to test the relationships among continuous variables, and the residuals were examined to ensure that they had an approximately normal distribution. All of the analyses were carried out with the Statistical Package for the Social Sciences, version 18.0 (SPSS Inc., Chicago, IL, USA). The threshold for statistical significance was set at p < 0.05.

Results

Table 1 lists the characteristics of the 40 patients included in the study, i.e., 16 males and 24 females, the mean age being 48 years (range, 26-83 years). Of the 40 patients, 11 were Black. Most of the 25 patients with pulmonary sarcoidosis showed a restrictive pattern or an
obstructive pattern, and 15 were current or former smokers. Twenty-seven patients received corticosteroid therapy.

Histological examination revealed noncaseating granulomas (Figure 1), composed of multinucleated giant cells, epithelioid cells, macrophages, and lymphocytes, in all of the patients. Staining for fungi and AFB gave negative results. In all of the patients with extrapulmonary sarcoidosis, the granulomas were characterized by severe architectural remodeling and variable amounts of fibrous tissue. In particular, the cortical and

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**Figure 1** - Lung, lymph node, liver, and skin tissue samples stained with H&E, picrosirius red (collagen), and Weigert’s resorcin-fuchsin (elastic fibers). Note architectural distortion in the lung, lymph node, liver, and skin samples (H&E staining), a diffuse increase in the birefringence of collagen fibers (picrosirius red staining), and a low amount of elastic fibers in the granulomas (Weigert’s resorcin-fuchsin stain). In decreasing order, similar distortions are observed in the lung, lymph node, liver, and skin tissue samples (magnification, x200).
Immunophenotyping and extracellular matrix remodeling in pulmonary and extrapulmonary sarcoidosis

Table 2 - Summary of morphometric results.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Lung</th>
<th>Lymph node</th>
<th>Liver</th>
<th>Skin</th>
<th>Extrapulmonary (total)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD4</td>
<td>7.48</td>
<td>16.89</td>
<td>10.08</td>
<td>8.18</td>
<td>11.71</td>
</tr>
<tr>
<td>CD8</td>
<td>5.03</td>
<td>14.92</td>
<td>12.10</td>
<td>4.57</td>
<td>10.53</td>
</tr>
<tr>
<td>CD3</td>
<td>7.73</td>
<td>20.36</td>
<td>11.81</td>
<td>12.37</td>
<td>14.84</td>
</tr>
<tr>
<td>CD20</td>
<td>2.75</td>
<td>11.38</td>
<td>1.51</td>
<td>2.72</td>
<td>5.20</td>
</tr>
<tr>
<td>CD1a</td>
<td>0.36</td>
<td>0.28</td>
<td>0.10</td>
<td>1.95</td>
<td>0.77</td>
</tr>
<tr>
<td>S100</td>
<td>1.10</td>
<td>1.83</td>
<td>1.71</td>
<td>1.68</td>
<td>1.74</td>
</tr>
<tr>
<td>CD68</td>
<td>9.64</td>
<td>19.22</td>
<td>12.08</td>
<td>11.27</td>
<td>14.19</td>
</tr>
<tr>
<td>Elastase</td>
<td>0.40</td>
<td>1.30</td>
<td>2.71</td>
<td>1.35</td>
<td>1.78</td>
</tr>
<tr>
<td>Collagen</td>
<td>25.62</td>
<td>15.08</td>
<td>6.30</td>
<td>5.15</td>
<td>8.84</td>
</tr>
<tr>
<td>Elastic fiber</td>
<td>3.75</td>
<td>48.39</td>
<td>49.82</td>
<td>29.79</td>
<td>42.66</td>
</tr>
</tbody>
</table>

The units of "% of points" indicate the number of points overlying the phenomenon of interest divided by the total number of points overlying the tumor. In morphometry, this is called a point fraction and is often symbolized as Pp, which has been shown to approximate the volume fraction.21

Figure 1 shows granulomas stained with picrosirius and viewed under polarized light. The granulomas in the samples obtained from the patients with extrapulmonary sarcoidosis showed a homogeneous orange-red birefringence, coincident with minor distortions of the ECM architecture. In contrast, the granulomas in the lung samples obtained from the patients with pulmonary sarcoidosis showed distortions of the ECM architecture, as well as a strong and heterogeneous birefringence. Large, sparse, and fragmented bundles of elastic fibers were more commonly observed in the granulomas in the extrapulmonary samples than in those in the lung samples. This correlates with the epithelioid and sclerosing areas seen in H&E preparations.

Figure 2 shows the immune cells in the pulmonary and extrapulmonary granulomas stained by immunohistochemistry; the most common cells around the granulomas found in the patients with extrapulmonary sarcoidosis were as follows: CD3+, CD4+, and CD8+ T-lineage cells; CD20+ lymphocytes; elastase-positive neutrophils; and CD68+ macrophages.

Table 2 shows the quantification of immune cells in the pulmonary and extrapulmonary granulomas. The morphological distribution of the immune cells in the granulomas coincided with the differences in quantification between the two forms of sarcoidosis. Overall, the extrapulmonary sarcoidosis samples showed a significantly higher density of CD4+ lymphocytes (p = 0.001), CD8+ lymphocytes (p = 0.001), CD3+ lymphocytes (p = 0.001), CD68+ lymphocytes (p = 0.03), and elastase-positive neutrophils (p = 0.001) surrounding or permeating granulomatous structures than did the pulmonary sarcoidosis samples. The number of CD4+ lymphocytes was significantly higher in the granulomas in the lymph node samples than in those in the liver (p = 0.05), skin (p = 0.03), and lung samples (p = 0.001). In addition, the number of CD8+ lymphocytes surrounding granulomas was significantly higher in lymph node samples than in lung and skin samples (p = 0.001 and p = 0.003, respectively). Furthermore, the number of CD8+ lymphocytes was significantly higher in liver samples than in lung samples (p = 0.01). Granulomas in the lymph nodes also showed a higher number of CD3+, CD20+, and CD68+ lymphocytes than did those in the lungs (p = 0.001, p = 0.01, and p = 0.004, respectively). The number of CD1a+ cells was higher in the granulomas found in the skin than in those found in the lungs, lymph nodes, and liver (p = 0.001 for all). The number of elastase-positive neutrophils permeating granulomas was higher in the liver than in the lungs (p = 0.02).
Figure 2 - Microscopic images of CD3+, CD4+, and CD8+ T lymphocytes; CD20+ B cells; CD68+ macrophages; CD1a+ dendritic cells; S100+ dendritic cells; and elastase-positive neutrophils in samples of lung, lymph node, liver, and skin parenchyma. It seems that CD4+ and CD8+ T lymphocytes are the most common cells infiltrating the lymph nodes, liver, skin, and lung. The lymph nodes also showed a significant number of CD3+, CD20+, and CD68+ cells. CD1a+ cells were most common in the skin, whereas elastase-positive neutrophils were prominent in the liver. The number of S100+ cells was similar in all the organs (immunohistochemical staining; magnification, x400).
The quantity of S100+ cells was very similar in the granulomas in the various organs studied. Positive associations were found between CD4+ and CD8+ cells \( (r = 0.55; p = 0.001) \), between CD8+ cells and elastase-positive neutrophils \( (r = 0.34; p = 0.03) \), between CD8+ cells and elastic fibers \( (r = 0.41; p = 0.01) \), between CD3+ cells and elastic fibers \( (r = 0.46; p = 0.004) \), between CD1a+ cells and collagen fibers \( (r = 0.35; p = 0.05) \), and between elastase-positive neutrophils and elastic fibers \( (r = 0.35; p = 0.03) \). Negative associations were found between CD8+ cells and collagen fibers \( (r = -0.46; p = 0.04) \), as well as between CD1a+ cells and elastic fibers \( (r = -0.34; p = 0.05) \).

The cellular immune components and reparative components correlated with clinical, radiological, and pulmonary function findings in the patients with pulmonary sarcoidosis. We found that CD4+ lymphocytes were negatively associated with smoking history \( (r = -0.73; p = 0.03) \), radiological opacities \( (r = -0.59; p = 0.04) \), and micronodules \( (r = -0.54; p = 0.04) \). Mediastinal lymph node granulomas were positively associated with CD8+ lymphocytes \( (r = 0.70; p = 0.004) \). An FVC of 70% of the predicted value was positively associated with CD68+ macrophages \( (p < 0.05) \). The FEV/FVC ratio was positively associated with CD4+, CD8+, and CD3+ lymphocytes. A TLC of 82% of the predicted value was negatively associated with CD1a+ cells \( (p < 0.05) \). Finally, there was a negative association between DLCO and collagen/elastic fibers \( (r = -0.90; p = 0.04) \).

**Discussion**

The pathological hallmark of sarcoidosis is the presence of noncaseating granulomas in various organs, including the lungs, lymph nodes, liver, skin, heart, and brain. In this chronic systemic disease, these histologically compact inflammatory lesions containing T lymphocytes and mononuclear phagocytes can appear and disappear insidiously in some patients, whereas, in others, the lesions can cause significant organ dysfunction due to persistent inflammation and obliteration of vital structures, resulting in extensive tissue fibrosis or cavitation. The most common and devastating effects occur in the pulmonary system. Therefore, the knowledge of ECM remodeling and immune response mechanisms in the pathogenesis of sarcoidosis, together with the early detection of the disease by methods that are more sensitive and aimed at therapy, will help improve long-term pulmonary function and will prevent the cumulative morbidity associated with the currently available therapy. The question of interest is whether further information can help us define the immune response and tissue remodeling in sarcoidosis on the basis of the histological characteristics and patterns of the granulomas, which will show a better correlation with pulmonary function tests and the natural history of the disease, as well as with the response to therapy.

The pathological process in the granulomatous involvement in sarcoidosis undoubtedly includes complex serial and sequential steps; among these, severe immune dysfunction and tissue remodeling are thought to be important, given that both lead to scarring. The present study showed a higher degree of immunoreactivity to CD4, CD8, CD3, CD68, and neutrophils in extrapulmonary sarcoidosis than in pulmonary sarcoidosis. In addition, the number of CD4, CD8, CD3, and CD20 lymphocytes was significantly higher in the lymph node granulomas, whereas CD1a dendritic cells and neutrophils were more prominent in the skin and lung granulomas, respectively. It is known that CD4+ T lymphocytes play an essential role in creating the microenvironment for B lymphocyte activation and differentiation following antigen exposure. In the lungs, the fact that follicular B lymphocyte aggregates are located almost exclusively in peribronchiolar areas beneath the epithelium of the bronchial branches suggests that they represent the development of bronchus-associated lymphoid tissue. These structures have been shown to be capable of mounting competent adaptive immune responses. Unlike typical lymph nodes, they are not dependent on afferent lymphatics for antigen retrieval; rather, they sample antigen directly from the lung lumen. This raises the question of whether an external factor could promote the formation of lymphoid follicles in sarcoidosis, thereby inducing or modulating the disease process. This question is especially interesting in light of the reported association between smoking and increased formation of peribronchiolar B cell follicles in some pulmonary diseases. Although CD4+ and CD8+ T lymphocytes are usually present in very low numbers in peripheral blood, increased numbers have been reported in apparently healthy individuals, as well as in various clinical conditions. In a recent study,
Moller et al. reported finding dominant Th1 cytokine expression, with elevated mRNA and protein levels of IL-12 and IFN-γ, but not IL-4, in sarcoid lung cells when compared with those in normal samples, supporting the notion of an exaggerated immunological/inflammatory response.

Consistent with these findings, various other immunological abnormalities have been identified, including tissue accumulation of CD4+ cells with helper-inducer activity, increased in situ production of cell-derived cytokines, B cell hyperactivity with spontaneous production of immunoglobulins, and accumulation of activated monocytic cells. These cells are known to play key roles in active tissue remodeling by releasing soluble mediators, such as cytokines and chemokines with proinflammatory and fibrogenic activity, reactive oxygen species, and proteolytic enzymes that have the capacity to degrade the connective tissue scaffold.

In our study, the granulomas in the samples obtained from patients with extrapulmonary sarcoidosis (lymph node, liver, and skin involvement) were found to be the prototype of the apposition of elastic fibers in the tissues. In contrast, the granulomas in the lung samples from patients with pulmonary sarcoidosis were found to represent the prototype of an increased number of collagen fibers and a lower degree of elastic apposition, characterizing a fibroelastotic process. The elastic system plays an important role in maintaining organ patency and the elastic recoil; therefore, elastosis might be important for the understanding of organ function.

Similar findings were reported by our group when studying the ECM in lung biopsy specimens from patients with idiopathic interstitial pneumonia. We hypothesized that elastosis is related to the inflammatory elastolysis observed at the initial stage of inflammation, reinforcing the notion that a proinflammatory mechanism is present in granulomatous diseases. The proinflammatory hypothesis is defended by various authors as a transient form of organizing fibrosis, and some of our findings in the present study seem to support this theory, given that the highest levels of neutrophils were observed in the patients with pulmonary sarcoidosis.

For all of these reasons, it would not be surprising to learn that cell immunostaining and the determination of collagen/elastic fiber density can provide relevant information on tissue remodeling in sarcoidosis. Our results confirm the pathogenetic implications of immune cell infiltration and the remodeling state of granulomas in pulmonary and extrapulmonary sarcoidosis. Although only a few studies focusing on the production of matrix proteins, TGF-β, fibrin, and matrix metalloproteinases were able to demonstrate a significant association between tissue remodeling and immune response in sarcoidosis, our results suggest that a fibroelastotic process, together with immune cell infiltration, probably contributes to tissue changes in sarcoidosis. More interestingly, our results suggest that antigen clearance is less efficient in pulmonary sarcoidosis than in extrapulmonary sarcoidosis. More specifically, we found that different functional tests corroborated the remodeling and immune cell regulation in pulmonary sarcoidosis, which is suggestive of a reparative effect on the functional and structural disarray found in the lung parenchyma.

The major limitations of the present study are related to the difficulty in comparing our results with those of other studies, given that there are few studies in the literature showing associations of immunophenotyping and tissue remodeling with standard clinical parameters (age, gender, smoking history, and duration of symptoms prior to lung biopsy), physiological tests, and dyspnea scores in sarcoidosis patients.

In summary, we have demonstrated the pathophysiological features of tissue remodeling and immune response in sarcoidosis. Particularly in pulmonary sarcoidosis, there are two different immune response and tissue remodeling pathways, with an impact on physiological tests. The immune and fibroelastotic processes observed in the parenchyma of the affected organs appear to be part of the general processes of inflammation and collagen/elastic fiber deposition, which have an independent course in sarcoidosis. Our findings suggest that there are two mechanisms of injury and repair in sarcoidosis, direct airway injury being more likely to occur in pulmonary sarcoidosis. Regardless of the pathogenetic determinants, the pathway involvement that is likely to progress to uncontrolled fibrosis and therefore shorten the life of patients with sarcoidosis should be identified so that the treatment is effective.

References


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